## Phenolic Glycosides from the Leaves of *Alangium platanifolium* var. *platanifolium*

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## Received March 8, 2000

Chemical investigation of *Alangium platanifolium* var. *platanifolium* has resulted in the isolation of nine phenolic glycosides that were identified by means of 1D and 2D NMR experiments. Among them, catechol and salicinol *O*- and 1-*O*- $\beta$ -D-(6-*O*- $\beta$ -D-apiofuranosyl)glucopyranosides, respectively **1** and **2**, and two compounds characterized as adducts of 2,6-dihydroxybenzoic acid with salicin (plataplatanoside, **3**) and 4-hydroxysalicin (4-hydroxyalangifolioside, **4**) were determined structurally as new compounds.

In a previous paper, the isolation was reported of three new megastigmane diglycosides, alangionosides A–C, from the leaves of *Alangium platanifolium* (Sieb. et Zucc.) Harms var. *platanifolium* (Alangiaceae), endemic to the western area of Japan.<sup>1</sup> Subsequent chemical investigation has afforded five known and four new phenolic glycosides (1-4). This paper deals with the structure elucidation of these new compounds.



1 R<sup>1</sup>=OH, R<sup>2</sup>=Glc*p*(6'-1")Api*f* 2 R<sup>1</sup>=CH<sub>2</sub>OH, R<sup>2</sup>=Glc*p*(6'-1")Api*f* 



The air-dried plant material was extracted exhaustively with MeOH. The extract was defatted, and then extracted successively with ethyl acetate and *n*-BuOH. The dried *n*-BuOH-soluble fraction was separated by means of various chromatographic methods to yield nine phenolic glycosides. In addition to the new compounds, **1**–**4**, five known compounds isolated were identified spectroscopically as salicin,<sup>2</sup> henryoside,<sup>2,3</sup> henryoside 6-*O*- $\beta$ -D-glucopyranoside,<sup>4</sup> alangifolioside,<sup>2</sup> and demethylalangiside.<sup>5</sup>

Compound **1** was isolated as an amorphous powder, with a composition of  $C_{17}H_{24}O_{11}$ , as determined by negative-ion HRFABMS. The IR and UV spectra indicated that **1** was a phenolic glycoside. The <sup>1</sup>H NMR spectrum showed the presence of four aromatic protons ( $\delta_C$  6.79, 6.84, 6.91, and 7.18) and two characteristic anomeric protons [ $\delta_H$  4.79 (d, J = 8 Hz) and 5.00 (d, J = 2 Hz)]. The <sup>13</sup>C NMR spectrum showed five signals for a terminal  $\beta$ -apiofuranoside unit,

**Table 1.** <sup>13</sup>C NMR Data for Compounds 1-4 (CD<sub>3</sub>OD, 100 MHz)

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carbon	1	2	3	4
1	148.6	157.2	156.2	150.4
2	146.9	132.4	129.0	133.5
3	117.3	129.9	129.8	117.7
4	125.0	124.0	123.7	153.6
5	121.2	130.1	129.5	114.3
6	119.4	117.6	116.6	118.6
7		61.1	67.3	29.7
1′	104.6	103.7	111.8	101.1
2'	75.0	75.1	161.1	160.6
3′	77.7	78.1	105.7	127.7
4'	71.7	71.7	132.5	138.7
5'	77.2	77.2	110.4	107.8
6'	68.8	68.9	163.5	159.9
7′			174.8	173.4
1″	111.2	111.2	103.1	104.2
2″	78.2	78.2	75.0	75.2
3″	80.6	80.5	78.2	78.3
4‴	75.1	75.1	71.5	71.6
5″	65.8	65.7	78.1	78.1
6″			62.6	62.7

six signals for a 6-substituted  $\beta$ -glucopyranoside unit,<sup>6</sup> and six signals for an aromatic ring, two of which bear hydroxyl substituents. From this evidence, the structure of **1** was elucidated as catechol *O*- $\beta$ -D-(6'-*O*- $\beta$ -D-apiofuranosyl)glucopyranoside.

Compound **2** was isolated as an amorphous powder, whose elemental composition was determined to be  $C_{18}H_{26}O_{11}$  in a similar manner to **1**. The <sup>1</sup>H and <sup>13</sup>C NMR spectra showed that the aglycon portion was salicinol<sup>2</sup> (Table 1), and the sugar moiety was the same as that of compound **1**. Therefore, the structure of **2** was determined to be the 6'-O- $\beta$ -D-apiofuranoside of salicin.

Plataplatanoside (**3**) was also isolated as an amorphous powder, and its elemental composition was determined to be  $C_{20}H_{22}O_{10}$ . The IR spectrum indicated the presence of hydroxyl groups (3384 cm<sup>-1</sup>) and aromatic ring(s) (1620, 1597, and 1462 cm<sup>-1</sup>) in the molecule, and the UV absorption maxima also denoted one aromatic ring or more. The <sup>13</sup>C NMR spectrum showed the presence of six signals for one terminal  $\beta$ -glucopyranose moiety, with the remaining 14 signals representing two aromatic rings, one carboxyl group, and one hydroxymethyl group. The <sup>1</sup>H NMR spectrum showed the presence of seven aromatic protons, and the <sup>1</sup>H–<sup>1</sup>H COSY spectrum confirmed the presence of two series of aromatic protons, one with three and the other

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with four. From these results, together with those of the <sup>13</sup>C NMR and HSQC experiments, compound 3 was presumed to be an ether adduct of 2,6-dihydroxybenzoic acid and salicin, like henryoside and alangifolioside. The linkage position was expected to be between one of the hydroxyl groups of 2,6-dihydroxybenzoic acid and that of salicin through an ether bond, because the 2,6-dihydroxybenzoic acid moiety was substituted asymmetrically, and the carbon signals of C-2 and C-7 were shifted by -3.2 ppm and +6.2 ppm compared with those of salicin<sup>2</sup> (Table 1). The arrangement of the substutuents of **3** was finally confirmed by a HMBC experiment in which H<sub>2</sub>-7a and -7b ( $\delta_{\rm H}$  5.19 and 5.34) showed cross-peaks with C-2' ( $\delta_{\rm C}$  161.1), C-1 ( $\delta_{C}$  156.2), C-2 ( $\delta_{C}$  129.0), and C-3 ( $\delta_{C}$  129.8), and the anomeric proton ( $\delta_{\rm H}$  4.95) correlated with C-1. Therefore, the structure of this isolate was elucidated as 3.

Compound 4 was isolated as an amorphous powder with an elemental composition of C<sub>20</sub>H<sub>22</sub>O<sub>11</sub>, which is one oxygen atom more than alangifolioside. The spectroscopic data indicated that **4** was similar to alangifolioside<sup>2</sup> (Table 1), and the <sup>1</sup>H NMR spectrum showed two protons [ $\delta_{\rm H}$  6.36 (d, J = 8 Hz) and 7.21 (d, J = 8 Hz)] on one of the aromatic rings and three protons coupled in an ABX system on the other aromatic ring. The <sup>13</sup>C NMR spectrum showed the presence of four aromatic carbon signals with hydroxyl substituents. From this evidence, compound 4 could be assigned as alangifolioside substituted with an additional hydroxyl group on the salicin moiety. The substitution pattern was confirmed by a HMBC experiment in which H-3 [ $\delta_{\rm H}$  6.51 (d, J = 3 Hz)] showed cross-peaks with C-7  $(\delta_{\rm C} 29.7)$  and the anomeric proton  $[\delta_{\rm H} 4.78 \text{ (d, } J = 8 \text{ Hz})]$ C-1 ( $\delta_{\rm C}$  150.4). Therefore, the structure of **4** was concluded to be 4-hydroxyalangifolioside.

From the biosynthetic point of view, it is interesting that three adducts of 2,6-dihydroxybenzoic acid and salicin, henryoside, alangifolioside, and plataplatanoside (**3**), were isolated together in this investigation. Henryoside is an ester of both building units, while alangifolioside and plataplatanoside (**3**) are probably the electrophilic adducts of the salicin cation (or corresponding saligenol cation) at the 3-position and the phenolic hydroxyl group of 2,6dihydroxybenzoic acid, respectively.

## **Experimental Section**

General Experimental Procedures. Melting points were determined with a Yanagimoto micromelting point apparatus and are uncorrected. Optical rotations were measured with a Union Giken PM-101 digital polarimeter. FTIR and UV spectra were recorded on a Shimadzu FTIR-4200 and Shimadzu UV-160A spectrophotometer, respectively. <sup>1</sup>H and <sup>13</sup>C NMR spectra were measured on a JEOL JNM  $\alpha$ -400 spectrometer at 400 and 100 MHz, respectively, with tetramethylsilane as internal standard. HRFABMS (negative-ion mode) were obtained with a JEOL JMS–SX-102 mass spectrometer with PEG-400 as the matrix.

Reversed-phase gravity column chromatography was performed on Cosmosil  $75C_{18}$ –OPN (Nakalai Tesque, Kyoto, Japan) ( $\Phi = 40$  mm, L = 25 cm) with a linear gradient solvent system [H<sub>2</sub>O–MeOH (9:1, 1 L)  $\rightarrow$  (1:1, 1 L); with fractions of 10 g being collected]. The droplet counter-current chromatograph (DCCC) (Tokyo Rikakikai, Tokyo, Japan) was equipped with 500 glass columns ( $\Phi =$ 2 mm, L = 40 cm). The ascending method was used with CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O–*n*-PrOH (9:12:8:2) as solvent, and 5-g fractions were collected and numbered according to the order of elution of the mobile phase. Preparative HPLC was performed on an Inertsil column (GL Science, Tokyo, Japan) (ODS,  $\Phi = 20$  mm, L = 25 cm), with the flow rate being 6 mL/min and detection being performed by UV at 254 nm.

**Extraction and Isolation.** The air-dried leaves of *A*. platanifolium var. platanifolium (Alangiaceae)<sup>1</sup> (3.55 kg) were extracted with MeOH (12 L  $\times$  2). To the concentrated MeOH extract (1.5 L) was added 75 mL of H<sub>2</sub>O, followed by washing with *n*-hexane (1.5 L). The MeOH layer was concentrated to a dark green mass and then suspended in 1.5 L of H<sub>2</sub>O. The suspension was extracted successively with EtOAc (1.5 L) and n-BuOH (1.5 L) to give dried residues of 15.8 and 65.9 g of EtOAc-soluble and *n*-BuOHsoluble fractions, respectively. The n-BuOH-soluble fraction was subjected to passage over a column of a highly porous synthetic resin (Diaion HP-20; Mitsubishi Kagaku, Tokyo, Japan), with 20% (6 L), 40% (6 L), 60% (6 L), and 80% (6 L) MeOH in H<sub>2</sub>O, and MeOH (6 L) as the eluents. Fractions of 2 L were collected. The residue (14.9 g) from the 40% MeOH eluate was subjected to Si gel (300 g) column chromatography by gradient elution with CHCl<sub>3</sub> and MeOH [CHCl<sub>3</sub> (1 L); CHCl<sub>3</sub>–MeOH (99:1, 2 L), (49:1, 2 L), (24:1, 4 L), (93:7, 4 L), (9:1, 4 L), (17:3, 4 L), (4:1, 4 L), (3:1, 4 L), and (7:3, 4 L); fractions of 500 mL were collected]. The residue (1.52 g in fractions 31-37) from the 7% MeOH eluate was then subjected to reversed-phase column chromatography (287 mg in fractions 105-120) and then to DCCC to give 28 mg of 2 in fractions 45-55.

The residue (2.01 g in fractions 59-80) of the 10-15%MeOH eluate obtained on Si gel column chromatography was subjected to reversed-phase column chromatography, and then the residue (65 mg in fractions 41-50) was purified by DCCC to give 25 mg of **4** in fractions 3-9. The residue (158 mg in fractions 61-70) was subjected DCCC to give 37 mg of henryoside 6'-*O*- $\beta$ -D-glucopyranoside in fractions 4-8 and 67 mg of crude **2** in fractions 16-22. The latter was finally purified by preparative HPLC [H<sub>2</sub>O-MeOH (17:3)] to give 42 mg of **2**. The residue (59 mg in fractions 71-76) was purified by DCCC to give 30 mg of **1** in fractions 21-26.

The 60% MeOH residue (9.20 g) from the 60% MeOH eluate obtained on Diaion HP-20 column chromatography was subjected to Si gel (250 g) column chromatography by gradient elution with CHCl<sub>3</sub> and MeOH [CHCl<sub>3</sub> (1 L), CHCl<sub>3</sub>–MeOH (99:1, 2 L), (49:1, 2 L), (24:1, 2 L), (47:3, 2 L), (23:2, 2 L), (9:1, 2 L), (7:1, 2 L), (17:3, 2 L), (4:1, 2 L), (3:1, 2 L), and (7:3, 2 L); fractions of 500 mL were collected]. The residue (772 mg in fractions 30-32) of the 8% MeOH eluate was successively separated by reversed-phase column chromatography (93 mg in fractions 122-127) and DCCC to give 28 mg of demethylalangiside in fractions 44-50.

The residue (10.6 g) from the 80% MeOH eluate obtained on Diaion HP-20 column chromatography was subjected to Si gel (300 g) column chromatography by gradient elution with CHCl3 and MeOH [CHCl3 (1 L), CHCl3-MeOH (99:1, 2 L), (49:1, 2 L), (24:1, 2 L), (47:3, 2 L), (23:2, 2 L), (9:1, 2 L), (7:1, 2 L), (17:3, 2 L), (4:1, 2 L), (3:1, 2 L), and (7:3, 2 L); fractions of 500 mL were collected]. The residue (1.46 g in fractions 24-32) of the 8% MeOH eluate was purified by reversed-phase column chromatography to give 119 mg of salicin in fractions 51-62. The residue (1.69 g in fractions 42-48) of the 12.5% MeOH eluate was subjected to reversed-phase column chromatography, and the resultant subfractions [fractions 59-70 (68 mg) and fraction 105–116 (61 mg)] were then purified by two separate DCCC runs to give 42 mg of 3 in fractions 8-12 and 43 mg of alangiside in fractions 12–18, respectively.

**Catechol** *O*-β-D-(6'-*O*-β-D-apiofuranosyl)glucopyranoside (1): amorphous powder;  $[\alpha]^{29}_{D} - 85.9^{\circ}$  (*c* 0.76, MeOH); IR (KBr)  $\nu_{max}$  3422, 2930, 1601, 1503, 1269, 1068, 754 cm<sup>-1</sup>; UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 204 (3.87), 274 (3.29) nm; <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$  3.59 (2H, s, H<sub>2</sub>-4"), 3.65 (1H, dd, *J* = 6, 11 Hz, H-6'a), 3.76 (1H, d, *J* = 10 Hz, H-5"a), 3.92 (1H, d, *J* = 2 Hz, H-2"), 3.98 (1H, d, *J* = 10 Hz, H-5"b), 4.02 (1H, dd, *J* = 2, 11 Hz, H-6'b), 4.70 (1H, d, *J* = 8 Hz, H-1'), 5.00 (1H, d, *J* = 2 Hz, H-1"), 6.79 (1H, ddd, *J* = 2, 7, 8 Hz, H-4), 6.84 (1H, dd, *J* = 2, 8 Hz, H-6), 6.91 (1H, ddd, *J* = 2, 7, 8 Hz, H-5), 7.18 (1H, dd, *J* = 2, 8 Hz, H-3); <sup>13</sup>C NMR (CD<sub>3</sub>OD), see Table 1; HRFABMS (negative-ion mode) *m*/*z* 403.1222 [M - H]<sup>-</sup> (calcd for C<sub>17</sub>H<sub>23</sub>O<sub>11</sub>, 403.1240).

**Salicin 6**'-*O*β-D-apiofuranoside (2): amorphous powder;  $[\alpha]^{29}_D - 61.6^{\circ}$  (*c* 0.76, MeOH); IR (KBr)  $\nu_{max}$  3407, 2928, 1601, 1493, 1458, 1237, 1068, 760 cm<sup>-1</sup>; UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 212 (3.77), 269 (3.88) nm; <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$  3.45 (1H, t, *J* = 9 Hz, H-3'), 3.50 (1H, dd, *J* = 8, 9 Hz, H-2'), 3.57 (2H, s, H<sub>2</sub>-4''), 3.63 (1H, dd, *J* = 7, 11 Hz, H-6'a), 3.74 (1H, d, *J* = 10 Hz, H-5''a), 3.90 (1H, d, *J* = 2 Hz, H-2'), 3.95 (1H, d, *J* = 10 Hz, H-5''b), 4.02 (1H, dd, *J* = 2, 11 Hz, H-6'b), 4.59 (1H, d, *J* = 13 Hz, H-7a), 4.76 (1H, d, *J* = 13 Hz, H-7b), 4.82 (1H, d, *J* = 8 Hz, H-1'), 4.98 (1H, d, *J* = 2 Hz, H-1''), 7.04 (1H, dt, *J* = 1, 8 Hz, H-4), 7.21 (1H, dd, *J* = 1, 8 Hz, H-6), 7.28 (1H, dt, *J* = 2, 8 Hz, H-5), 7.34 (1H, dd, *J* = 2, 8 Hz, H-3); HRFABMS (negative-ion mode) *m*/*z* 417.1407 [M - H]<sup>-</sup> (calcd for C<sub>18</sub>H<sub>25</sub>O<sub>11</sub>, 417.1379).

**Plataplatanoside (3):**  $[α]^{29}_D - 18.9^\circ$  (*c* 0.85, MeOH); IR (KBr)  $\nu_{max}$  3384, 2944, 1620, 1597, 1462, 1381, 1221, 1084, 1042, 754 cm<sup>-1</sup>; UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 212 (4.36), 248 (3.67), 274 (3.20), 303 (3.48) nm; <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$  3.72 (1H, dd, *J* = 5, 12 Hz, H-6"a), 3.89 (1H, dd, *J* = 2, 12 Hz, H-6"b), 4.95 (1H, d, *J* = 8 Hz, H-1"), 5.19 (1H, d, *J* = 14 Hz, H-7a), 5.34 (1H, d, *J* = 14 Hz, H-7b), 6.42 (1H, dd, *J* = 1, 8 Hz, H-5'), 6.48 (1H, dd, *J* = 1, 8 Hz, H-3'), 7.01 (1H, dt, *J* = 2, 8 Hz, H-4), 7.07 (1H, t, *J* = 8 Hz, H-4'), 7.19 (1H, dd, *J* = 2, 8 Hz, H-6), 7.23 (1H, dt, *J* = 2, 8 Hz, H-5), 7.64 (1H, dd, *J* = 2, 8 Hz, H-3); HRFABMS (negative-ion mode) *m*/*z* 421.1147 [M - H]<sup>-</sup> (calcd for C<sub>20</sub>H<sub>21</sub>O<sub>10</sub>, 403.1134). **4-Hydroxyalangifolioside (4):**  $[\alpha]^{29}{}_{\rm D}$  -7.1° (*c* 1.40, MeOH); IR (KBr)  $\nu_{\rm max}$  3420, 1674, 1626, 1490, 1449, 1344, 1213, 1074, 1040, 814 cm<sup>-1</sup>; UV (MeOH)  $\lambda_{\rm max}$  (log  $\epsilon$ ) 208 (4.30), 269 (3.17), 292 (3.47), 310 (3.44) nm; <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$  3.61 (1H, dd, J = 5, 12 Hz, H-6"a), 3.70 (1H, dd, J = 2, 12 Hz, H-6"b), 3.90 (1H, d, J = 16 Hz, H-7a), 3.96 (1H, d, J = 16 Hz, H-7b), 4.78 (1H, d, J = 8 Hz, H-1"), 6.36 (1H, dd, J = 3 Hz, H-5'), 6.51 (1H, dd, J = 8 Hz, H-3), 6.56 (1H, dd, J = 8 Hz, H-5), 7.04 (1H, d, J = 8 Hz, H-6), 7.21 (1H, d, J = 8 Hz, H-4'); HRFABMS (negative-ion mode) m/z 437.1101 [M - H]<sup>-</sup> (calcd for C<sub>20</sub>H<sub>21</sub>O<sub>11</sub>, 403.1084).

**Known compounds isolated:** salicin, colorless crystals, mp 201–204 °C,  $[\alpha]^{31}_D$  –39.2° (*c* 0.74, MeOH); henryoside, colorless crystals, mp 133–135 °C,  $[\alpha]^{31}_D$  –36.7° (*c* 0.90, pyridine); henryoside 6′-*O*-*β*-D-glucopyranoside, amorphous powder,  $[\alpha]^{23}_D$  –36.9° (*c* 2.52, MeOH); alangifolioside, colorless needles, mp 125–128 °C,  $[\alpha]^{31}_D$  –16.9° (*c* 0.83, pyridine); demethylalangiside, colorless crystals, mp 180–182 °C,  $[\alpha]^{31}_D$  –72.6° (*c* 0.84, MeOH).

Acknowledgment. The authors are grateful for the access to the superconducting NMR instrument in the Analytical Center of Molecular Medicine of Hiroshima University Faculty of Medicine.

## **References and Notes**

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NP000119L